

# Non-denaturing extraction for ELISA and dipstick activity assays

## Introduction

A critical step in performing any biological assay is preparing the test sample. Antibody-based ELISA and Dipstick assays capture sample proteins and protein complexes in their native, non-denatured state. Many assays, notably our OXPHOS assays, capture membrane-bound multi-subunit enzymes. Sample proteins for these assays must be extracted by non-denaturing methods, not just for recognition by the capture antibodies, but also to retain their enzymatic function for activity assays. Therefore, we have developed assays using a non-denaturing detergent, n-Dodecyl-beta-D-maltopyranoside (CAS# 69227-93-6), which is commonly referred to as lauryl maltoside. Lauryl maltoside is a non-ionic detergent used for the stabilization and activation of enzymes in membrane research, and thus is well suited for the analysis of mitochondrial membrane proteins and enzyme complexes 1-7.

The guideline outlines the steps to perform lauryl maltoside extractions of functional enzymes and proteins from tissues and cultured cells for use with our ELISA and Dipstick assays.

## Tissue preparations

Many of our assays are reactive with rodent proteins and were developed with rodent tissues (mouse and rat). The data in this section of the guide was obtained with rodent tissues, but the procedure applies to human and other animal tissues as well. Fully functional enzymes can be extracted from tissues by following a few simple steps.

We use simple mechanical disruption with Dounce homogenizers to disrupt cells minimally. Tissue is homogenized sequentially with two different-sized pestles and processed until smooth enough to be pipetted. Sequential homogenization is started with a large-clearance pestle that provides a finer disruption of the tissue. A suitable Dounce homogenizer is available from us in our tissue sample preparation kit, ab110169.

For long-term storage, it is best to make tissue homogenates that are as concentrated as possible. We have found that tissue can be homogenized with Dounce homogenizers with relative ease up to 25 mg/ml. After homogenization, sample detergent lysates can be made immediately or tissue homogenates can be aliquoted and frozen at -80°C.

Tissue	Wet weight (mg/ml)	Buffer expected protein concentration (by BCA)
Brain	500	25 mg/ml
Heart	375	25 mg/ml
Kidney	375	25 mg/ml
Liver	300	25 mg/ml

### **Table 1: Typical wet weight starting guide**

*The amount of tissue homogenized per ml of buffer to make a homogenate that is approximately 25 mg/ml protein by BCA protein analysis.*

## Materials for tissue homogenization

- Dounce homogenizer, 2 ml (Kontes, 885300-0002 or ab110169)
- Homogenization buffer (PBS)
- Protease inhibitor cocktail (ab65621)
- BCA™ protein assay
- Weighing balance
- Scalpel
- 2.0 ml microtubes

## Method for tissue homogenization

1. Add 1 ml buffer (containing protease inhibitor) to chilled mortar on ice.
2. Wash tissue to remove any blood and trim to remove any connective tissue and fat. Weigh and mince the appropriate amount of tissue and add it to the 1 ml of buffer in the mortar.
3. Perform 20-40 strokes with the large-clearance pestle A until smooth.
4. Perform 20-40 strokes with the small-clearance pestle B until smooth (pestles A and B are included with Kontes 885300-0002 and also ab110169).
5. Using a pipette, transfer homogenate to a 2.0 ml microtube on ice.
6. Perform a BCA™ protein assay to determine protein concentration.
7. Proceed to detergent extraction or aliquot and freeze the homogenate.

## Detergent extraction of proteins from tissue preparations

Protein extraction with lauryl maltoside is, in most cases, optimal with samples that contain a protein concentration of 5 mg/ml.

To extract protein from the prepared tissue homogenate, MitoSciences kits use one or two methods. We have noticed no significant difference in protein yield or assay performance between these two methods.

1. 1X extraction buffer (contains 1X lauryl maltoside detergent).
2. Add 1/10 volume of 10X lauryl maltoside detergent.

### Exceptions:

**ATP synthase** – For the Complex V microplate activity assay ab109704, it is recommended to thoroughly freeze/thaw the sample and wash the pelleted tissue homogenate to remove any soluble, non-membrane associated proteins. Concentrating the membrane proteins before protein assay leads to an improved detergent solubilization step and maximizes enzyme stability and oligomycin sensitivity.

**PDH** – For the PDH microplate activity assay ab109902, it is recommended to add the supplied detergent to a protein concentration of 25 mg/ml (undiluted homogenate) in order to maintain enzyme stability. Extracts should be centrifuged at a low speed (1,000 g).

## Materials for protein extraction

- 1X extraction buffer (1.5% lauryl maltoside)

or

- 10X lauryl maltoside detergent (15% lauryl maltoside solution)
- PBS

- Microtubes
- High-speed benchtop microcentrifuge
- BCA™ protein assay (ab102536)

or

- NanoDrop™ 2000 spectrophotometer (Thermo)

#### Protein extraction method I with 1X extraction buffer:

1. Resuspended tissue homogenate to 5 mg/ml in MitoSciences 1X extraction buffer (1.5% lauryl maltoside), e.g. dilute 100 µl of 25 mg/ml homogenate with 400 µl extraction buffer to a total volume of 500 µl. Add protease inhibitors.
2. Mix well and incubate on ice for 30 min.
3. Centrifuge at 16,000 x g for 20 min at 4°C.
4. Collect supernatant and discard the pellet.
5. Measure protein concentration with BCA™ assay or NanoDrop™ spectrophotometer.

#### Protein extraction method II with 10X lauryl maltoside:

1. Resuspend and dilute the cell pellet with 8 volumes of PBS (e.g. 50 µl pellet + 400 µl PBS to a total volume of 450 µl). Add protease inhibitors.
2. Add a 10-fold dilution of 10X lauryl maltoside (e.g. add 50 µl to the cell suspension for a total volume of 500 µl).
3. Mix well and incubate on ice for 30 min.
4. Centrifuge at 16,000 x g for 20 min at 4°C.
5. Collect supernatant and discard the pellet.
6. Measure protein concentration with BCA™ assay or NanoDrop™ spectrophotometer.

The protein concentration of the extract should now be  $\leq 5$  mg/ml, depending on extraction efficiency, as measured by BCA™ assay. To measure protein concentration with a NanoDrop™ spectrophotometer, follow the manufacturer's instructions for measuring protein and in the pull-down menu for sample type, select "other protein (E 1%)", then enter an extinction coefficient (Ext. Coedd, E 1% l/gm-cm) from table 2 below.

Cell line	Extinction coefficient
HepG2	8.1
HeLa	7.2
HL-60	8.6

**Table 2: Extinction coefficients for cell extracts**

***It may be possible to freeze the extract at -80°C before running the assay without significant loss of activity.***

## References

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