Map proteins/histone modifications to genomic loci



Cross-link

Sonication

Start with 2 X 150 cm² dishes of confluent cells (1 X 10² - 5 X 10² cells per dish)
Add formaldehyde to a final conc of 0.75% Incubate 2 - 30 min RT
Add glycine to a final conc of 125 mM
Shake gently for 5 min



Cross-linking fixes proteins to DNA.

Cell collection Wash cells with ice cold PBS

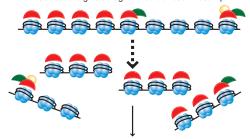
Scrape and collect cells into 5 ml ice cold PBS and transfer to a new tube

Wash dishes with 3 ml PBS to ensure all cells collected

Cell lysis

Centrifuge 5 min 1 000 g and remove supernatant
Add FA lysis buffer to cell pellet (750 µl per 1 X 10<sup>7</sup> cells)

Sonicate to give a fragment size of 500 - 1000 bp



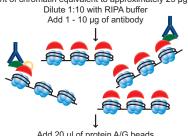
Sonication generates sheared, soluble chromatin. Optimize by performing a time course and purify DNA. Analyse fragment size on 1.5 % agarose gel.

Centrifuge 30 sec 4°C 8 000 g to pellet cell debris Transfer supernatant containing chromatin to a new tube

Remove 50  $\mu$ I (INPUT) and purify DNA to calculate the DNA concentration

Use an amount of chromatin equivalent to approximately 25 µg of DNA per IP Dilute 1:10 with RIPA buffer

Immunoprecipitation



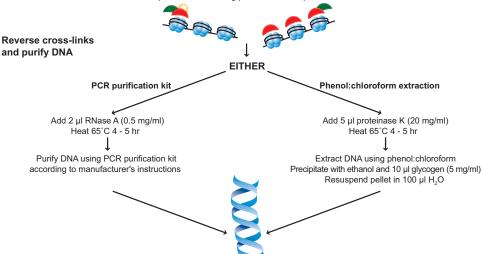
Antibody binds to target and associated DNA is isolated. DNA fragments not associated are removed during washes.

Add 20 µl of protein A/G beads Rotate overnight 4°C Centrifuge 1 min 2 000 g Wash 3 x with wash buffer Wash 1 x with final wash buffer.

Elute protein / DNA complex

Add 120 µl of Elution Buffer to the protein A/G beads Rotate 15 min 30°C

Centrifuge 1 min 2 000 g
Transfer supernatant containing protein/DNA complex to a new tube



DNA Analysis

Analyze DNA isolated by: quantitative PCR, sequencing (ChIP-seq), microarray (ChIP-chip).

